SYNTHESIS OF TRIANTENNARY BLOOD GROUP I ANTIGENS: *NEOLACTO-*GLYCOPENTADECAOSYL CERAMIDE¹

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Abstract: A stereocontrolled total synthesis of triantennary *neolacto*-glycopentadecaosyl ceramide was achieved for the first time.

Branched poly-N-acetyl-lactosamine type structures that are expressed on the cell surface of embryonic and transformed cells have significant biological functions in terms of cell-cell recognition and differentiation². Synthetic studies on the poly-N-acetyl-lactosamine type glycosyl ceramides have so far been focussed mainly for linear³ and biantennary⁴ *neolacto* series. Interested by the isolation and characterization of triantennary poly-N-acetyllactosamine type glycosyl ceramide 2 from rabbit erythrocyte membranes⁵ as well as of other related structures from human placenta⁶, we have studied a reasonable synthetic approach toward the triantennary structures 1 and 2. The retrosynthetic analysis shown in scheme 1 led us to design a glycosyl donor 3 and a ceramide derivative 4. The former may be further disconnected into glycosyl donors 5 and 6⁴ (or 7), and a glycohexaosyl acceptor 8. The key intermediate 8 may be reconstructed from glycosyl donor 9 and glycotetraosyl acceptor 10.



The glycohexaosyl acceptor 8 was synthesized as follows. The known compounds 11^4 was converted in two steps into fluoride 13^7 via 12^7 (*I* CAN⁸ in 4:3:2 CH₃CN-toluene-H₂O, 2 DAST⁹ in (C1CH₂)₂, 89% overall). Selective protection of diol 14^4 was carried out by treatment with LevOH, 2-chloro-1-methylpyridinium iodide (CMPI) and DABCO¹⁰ in (ClCH₂)₂ for 40 min at 25° to give nearly quantitative yield of 10^7 . Glycosylation of 10 with 13 in the presence of Cp₂HfCl₂¹¹,

AgOTf and powdered molecular sieve 4A (MS4A) in $(ClCH_2)_2$ for 3.5h at -23° afforded 96% of 15⁷. Three levuloyl groups in 15 was easily removed by NH₂NH₂•AcOH in 3:1 EtOH-THF to give 94% of the designed key intermediate triol 8⁷.



Having prepared compound 8, synthesis of glycododecaosyl ceramide 1 was first studied. $Cp_2Hf(OTf)_2$ promoted¹¹ coupling of 8 with 2.7 equivalents of 5 that was readily available from corresponding hemiacetal¹² by treatment with DAST afforded 92% of glycododecaoside 16⁷. Conversion of 16 into trichloroacetimidate 17 was achieved in 4 steps (1 10% Pd(OH)₂-C and H₂ in 16:1 MeOH-EtOAc, 2 Ac₂O and DMAP in Py at 45° for 18h, 3 NH₂NH₂•AcOH¹³ in DMF at 50° for 25min, 4 CCl₃CN¹⁴, DBU in (ClCH₂)₂ at -5° for 1h, 56% overall). TMSOTf promoted coupling of 17 with ceramide derivative 4¹⁵ in CHCl₃ at -23° afforded 45% of 18 which was further converted into 1 in 3 steps (1 40% MeNH₂¹⁶ in MeOH, 2 Ac₂O and DMAP in Py, 3 NaOMe in 7:4 MeOH-THF, 54% overall).



In studying the synthesis of 2 the known glycosyl donor 6 readily obtainable⁴ from 19 was first employed. Cp₂Hf(OTf)₂ promoted glycosylation of 8 with 2.7 equivalents of 6 in (ClCH₂)₂ at -23° afforded 90% of desired compound 21 which, however, could be converted into 23 only with difficulty mainly due to the sluggish hydrogenolytic removal of 33 benzyl groups in 21. In



order to decrease the number of benzyl groups in the pentadecaoside 21 we designed a completely acylated donor 7. Compound 19^4 was converted into 7^7 via 20^7 in 4 steps (1 10% Pd-C and H₂ in 10:5:1 MeOH-EtOAc-H₂O, 2 Ac₂O and DMAP in Py, 3 CAN in 50:38:25 CH₃CN-toluene-H₂O, 4 DAST in (ClCH₂)₂, 79% overall). With 2.1 equivalents of the glycotriosyl donor 7 Cp₂Hf(OTf)₂ promoted glycosylation of 8 in (ClCH₂)₂ at -23° gave 71% of 22, which was to our delight smoothly converted into 23 in two steps (1 10% Pd-C and H₂ in 16:1 MeOH-EtOAc, 2 Ac₂O and DMAP in Py, 60% overall). Conversion of 23 into 25 in 19% overall yield was carried out in 3 steps via trichloracetimidate 24 as described for 18. Finally, conversion of 25 into the target molecule 2 was executed in 3 steps as described for 1(1 40% MeNH₂ in MeOH for 3 days at 25°, 2 Ac₂O and DMAP in Py for 23h at 58°, 3 NaOMe in 7:4 MeOH-THF, 41% overall). The ¹H-nmr data for synthetic 1 and 2 were in good agreement with those for natural samples⁵, thus confirming the assigned structure for both 1 and 2.



In summary, by employing a key glycohexaosyl acceptor 8 and acetylated glycosyl donors 5 and 7, a versatile synthetic route to triantennary poly-N-acetyl-lactosamine type *neolacto*-glycosyl ceramide 1 and 2 was developed.

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- S. B. Levery, E. D. Nudelman, M. E. Salyan, and S. Hakomori, Biochemistry, 28 7772 (1989). 7 Physical data for new compounds are given below, values of $[\alpha]_D$ and $\delta_{H,C}$ were measured at 25°±3° for solutions in CHCl3 and CDCl3, respectively, unless noted otherwise. Signal assignment such as 1³ stands for a proton at C-1 of sugar residue 3. 1: RF 0.26 in 55:45:12 CHCl3-MeOH-H2O; [a]D -1.3° (c 0.1, py); 8H (49:1:1 DMSOd6-D2O-CD3OD, 60°) 0.845 (t, 6.4Hz, 2Me), 4.173 (d, 8.1Hz, 1¹), 4.233 (d, 7.0Hz, 18,8'.8"), 4.276 (d, 7.3Hz, 12), 4.306 (d, 6.6Hz, 14.6), 4.428 (d, 8.4Hz, 17',7"), 4.667 (d, 8.4Hz, 1^{3,5,7}), 5.364 (dd, 7.0 and 15.6Hz, 4^{Cer}), 5.557 (td, 7.0 and 15.0Hz, 5Cer). 2: RF 0.19 in 65:35:8 CHCl3-MeOH-H2O; 8H (49:1:1 DMSOd6-D2O-CD3OD, 60°) 0.858 (t, 7.0Hz, 2Me), 4.182 (d, 8.1Hz, 1¹), 4.286 (d, 7.3Hz, 1²), 4.315 (d, 7.3Hz, 1^{4,6,8,8',8"}), 4.438 (d, 8.4Hz, 1^{7',7"}), 4.690 (d. 8.4Hz, 1^{3,5,7}), 4.864 (d. 3.7Hz, 1^{9,9',9"}), 5.378 (dd, 7.0 and 15.4Hz, 4^{Cer}), 5.565 (td. 7.0 and 15.0Hz, 5Cer). 5: Rp 0.48 in 1:1 toluene-EtOAc; 8H 1.924, 1.969, 2.048, 2.074, 2.139, and 2.166 (6s, 6Ac), 6.108 (dd, 8.0 and 52.6Hz, 1¹). 7: R_F 0.41 in 2:3 toluene-EtOAc; δ_H 1.915, 1.947, 2.051, 2.057, 2.066, 2.134 x 2, 2.153, and 2.159 (8s, 9Ac), 5.779 (dd, 8.2 and 10.4Hz, 3¹), 6.102 (dd, 8.0 and 52.4Hz, 1¹). 8: R_F 0.36 in 1:1 toluene-EtOAc; [α]_D -1.4° (c 1.4); δ_H 1.069, 1.166, and 1.194 (3s, 3Piv), 1.623, 1.874, and 2.045 (3s, 3Ac), 5.232 (d, 3.7Hz, 4²), 5.391 (d, 8.4Hz, 1⁵). 10: R_F 0.39 in 1:1 hexane-EtOAc; [α]_D +2.4° (c 1.1); δ_H 1.079, 1.188, and 1.214 (3s, 3Piv), 1.685, 1.898, and 2.079 (3s, 3Ac), 2.173 (s, Lev), 5.112 (d, 8.4Hz, 1³), 5.333 (d, 3.7Hz, 4²). 12: RF 0.36 in 1:1 toluene-EtOAc, [a]D +46.9° (c 1.3); $\delta_{\rm H}$ 2.155 and 2.181 (2s, 2Lev), 5.285 (d, 0.9H, 8.4Hz, 1⁷B), 5.319 (d, 0.1H, 3.7Hz, 1⁷a). 13: RF 0.56 in 1:1 toluene-EtOAc; 8H 5.626 (dd, 0.15H, 2.6 and 54.2Hz, 1¹ a), 5.836 (dd, 0.85H, 7.7 and 53.9Hz, 1¹ β). 15: R_F 0.35 in 3:2 tolucne-EtOAc; [α]_D +2.8° (c 1.0); δ_H 1.068, 1.165, and 1.194 (3s, 3Piv), 1.628, 1.871, and 2.026 (3s, 3Ac), 2.121, 2.126, and 2.153 (3s, 3Lev), 4.928 (d, 8.4Hz, 1³), 5.225 (d, 3.7Hz, 4^2), 5.386 (d, 8.4Hz, 1^5). 16: R_F 0.28 in 2:3 toluene-EtOAc; [α]_D -13.6° (c 0.9); $\delta_{\rm H}$ 1.065, 1.159, and 1.179 (3s, 3Piv), 1.754, 1.864, 1.872, 1.885, 1.889, 1.897, 1.960, 1.963, 1.984, 2.017, 2.020, 2.032 x 3, 2.047, 2.052, 2.131, 2.137 x 2, 2.153, and 2.170 (18s, 21Ac), 5.708-5.634 (3dd, 3⁵, 3⁷, and 37'). 17: RF 0.45 in 1:3 toluene-EtOAc; 8H 1.088, 1.190, and 1.209 (3s, 3Piv), 6.440 (d, 3.3Hz, 1¹), 7.70-7.90 (m, 5Phth), 8.588 (s, NH). 18: RF 0.58 in 1:3 toluene-EtOAc; $[\alpha]_D$ +9.4° (c 0.5); δ_H 0.878 (t, 7Hz, 2Me), 1.097, 1.127, and 1.253 (3s, 3Piv), 1.676-2.172 (28s, 29Ac), 5.831 (td, 6.6 and 15.0Hz, 5^{Cer}), 7.35-8.00 (m, 5Phth and Bz). 20: RF 0.23 in 1:1 toluene-EtOAc; δ_H 1.923-2.156 (9s, 9Ac), 3.725 (s, OMe), 5.833 (d, 8.4Hz, 1¹). 21: RF 0.44 in 3:1 toluene, EtOAc; $[\alpha]_D$ +13.6° (c 1.2); δ_{H} 1.067, 1.166, and 1.180 (3s, 3Piv), 1.626, 1.824, 1.866, 1.874, 1.903, and 2.014 (6s, 6Ac). 22: RF 0.36 in 2:5 toluene-EtOAc; $[\alpha]_D$ +20.1° (c 1.3); δ_H 1.066, 1.162, and 1.180 (3s, 3Piv), 1.767-2.221 (23s, 30Ac). 23: RF 0.23 and 0.25 in 1:3 toluene-EtOAc; 5H 6.227 (d, 3.7Hz, 1¹), 7.60-7.95 (m, 5Phth). 24: RF 0.46 in 1:4 toluene-EtOAc; 8H 1.088, 1.190, and 1.208 (3s, 3Piv), 6.439 (d, 3.7Hz, 1¹), 7.65-7.95 (m, 5Phth), 8.590 (s, NH). 25: R_F 0.55 in 1:4 toluene-EtOAc; $[\alpha]_D$ +38.9° (c 0.3); δ_H 0.878 (t, 6.8Hz, 2Me), 1.097, 1.128 and 1.216 (3s, 3Piv), 1.679-2.177 (24s, 38Ac), 5.832 (td, 7.3 and 15.0Hz, 5^{Cer}), 7.30-7.98 (m, SPhih and Bz).
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